

Novel Technique Explains Herbicide's Link to Parkinson's Disease

BY [WILL DOSS](#) ON OCT 23, 2017

Northwestern Medicine scientists have used an innovative gene editing technique to identify the genes that may lead to Parkinson's disease after exposure to paraquat, a commonly-used herbicide.

This study, which utilized the CRISPR-Cas9 gene-editing tool, serves as a proof-of-concept for using genetic screens to investigate the biology of oxidative stress, according to senior author [Navdeep Chandel, PhD](#), the David W. Cugell, MD, Professor of [Medicine](#) in the Division of [Pulmonary and Critical Care](#) and of [Cell and Molecular Biology](#).

The study was [published](#) in *Nature Chemical Biology* and the first author was Colleen Reczek, PhD, a postdoctoral fellow in [Chandel's lab](#). Other authors included Chandel lab members Hyewon Kong, a student in the Walter S. and Lucienne Driskill Graduate Program in Life Sciences, and Inmaculada Martinez-Reyes, PhD, a postdoctoral fellow.

The use of paraquat, which causes cell death via oxidative stress, is restricted in the United States and banned in the European Union, but the chemical is still used widely throughout Asia and the developing world, according to Chandel. Ingestion of paraquat can lead to lung fibrosis or even death, but a [2011 study](#) linked occupational use to an increased risk for Parkinson's disease, renewing interest in its impact on humans.

A major cause of Parkinson's is the loss of function in dopamine neurons in a small brain region called the substantia nigra pars compacta, according to previous research. Those neurons are known to be highly vulnerable to oxidative stress, leading scientists to hypothesize paraquat was linked to Parkinson's disease through this oxidative stress.

"Paraquat generates a lot of oxidants. Naturally those dopaminergic neurons will be the most susceptible to damage," said Chandel, who is also a member of the [Robert H. Lurie Comprehensive Cancer Center of Northwestern University](#).

However, the mechanism by which paraquat created oxidants was unknown — until now.

Chandel and his collaborators conducted a CRISPR-Cas9 positive-selection screen, creating thousands of cells, each with one individual gene turned off.

"We thought it was a metabolic protein that paraquat was activating to generate oxidants," Chandel said. "So we localized our work to the 3,000 genes that encode for metabolic proteins, rather than the 18,000 to 20,000 genes human cells have in total."

They exposed that subset of cells to paraquat – the majority of cells died, but not all of them. Certain cells with knocked-out genes were resistant to paraquat, suggesting those genes may be responsible for the toxicity.

Scientists identified three genes whose loss conferred resistance to paraquat: POR, ATP7A and SLC45A4. POR, a protein in the endoplasmic reticulum, was fingered as the main source of oxidation that caused the damage. Pinpointing these genes could help identify people who are especially vulnerable to paraquat, Chandel said.

"Certain people with genetic mutations could have high levels of this gene. They would be very susceptible to paraquat poisoning while working on a farm, for example," he said.

However, the most impactful takeaway from the paper may be as a proof-of-concept for investigative biology of oxidative stress, according to Chandel.

POR had been previously implicated in oxidant generation, but the majority of evidence had pointed to systems in the mitochondria, according to the study, and no definitive answer had emerged until this study was conducted.

"Now, we can go in and test how agents of oxidant stress work," Chandel said. "The beauty of the paper is in the power of these unbiased genetic screens we can now use with CRISPR technology."

Investigating oxidant stress could pay dividends in the future, according to Chandel, including in the development of drugs designed to generate oxidative stress in cancer cells, killing them while leaving healthy cells alone. While some drugs currently exist, not enough is known about their pathways to create a functioning compound, Chandel said.

"The biology of oxidative stress is still a mystery," he said. "CRISPR positive-selection screens could be a way to figure it out."

The Northwestern Medicine investigators collaborated with co-authors David Sabatini, PhD, professor of biology at Massachusetts Institute of Technology (MIT) Department of Biology; Tim Wang, graduate student at MIT; and Kivanc Birsoy, PhD, Chapman Perelman Assistant Professor at The Rockefeller University.

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